

The MocR/GabR ectoine and hydroxyectoine catabolism regulator EnuR:

Inducer and DNA binding

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Running title: Genetic control of ectoine/hydroxyectoine utilization

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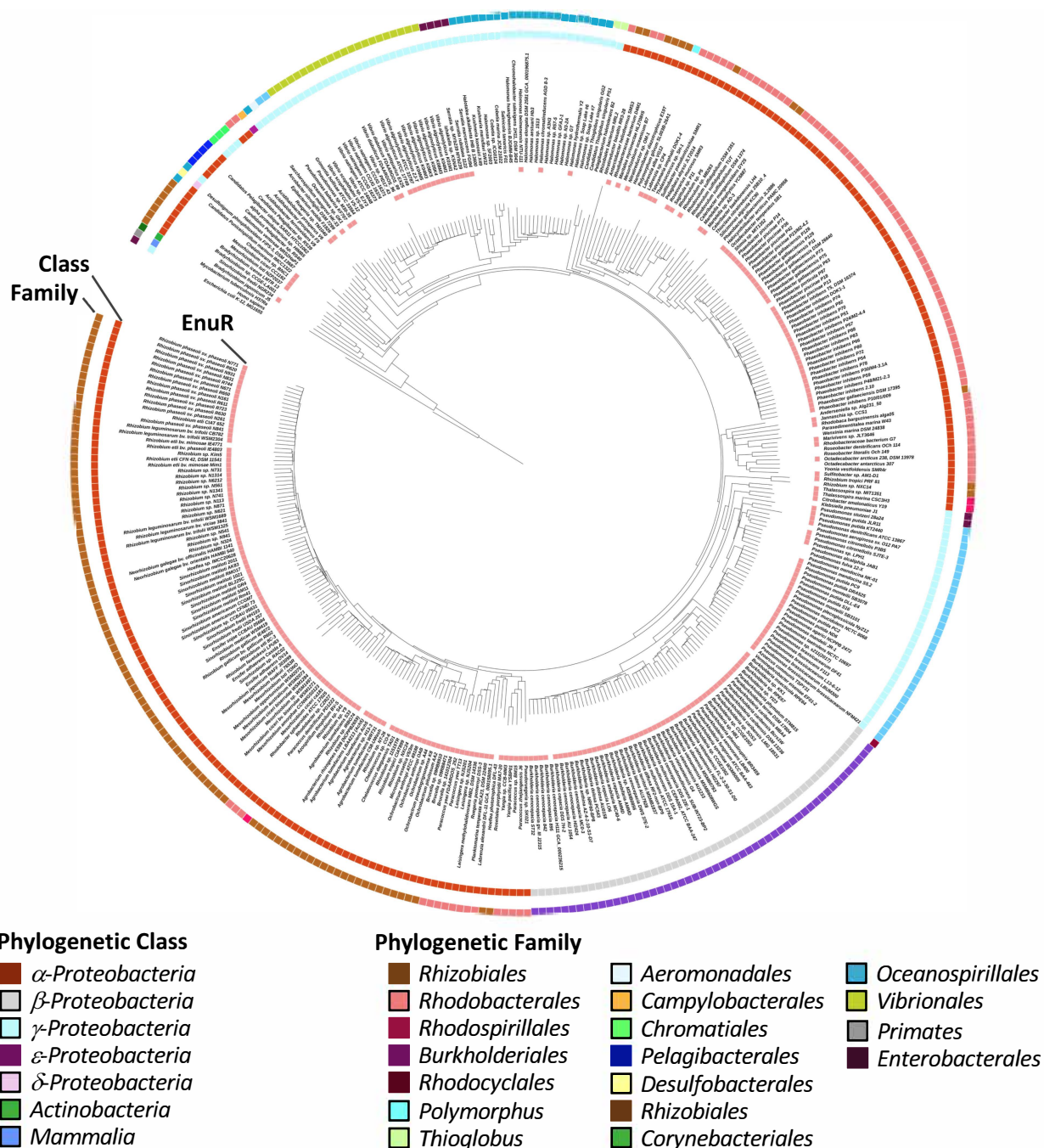
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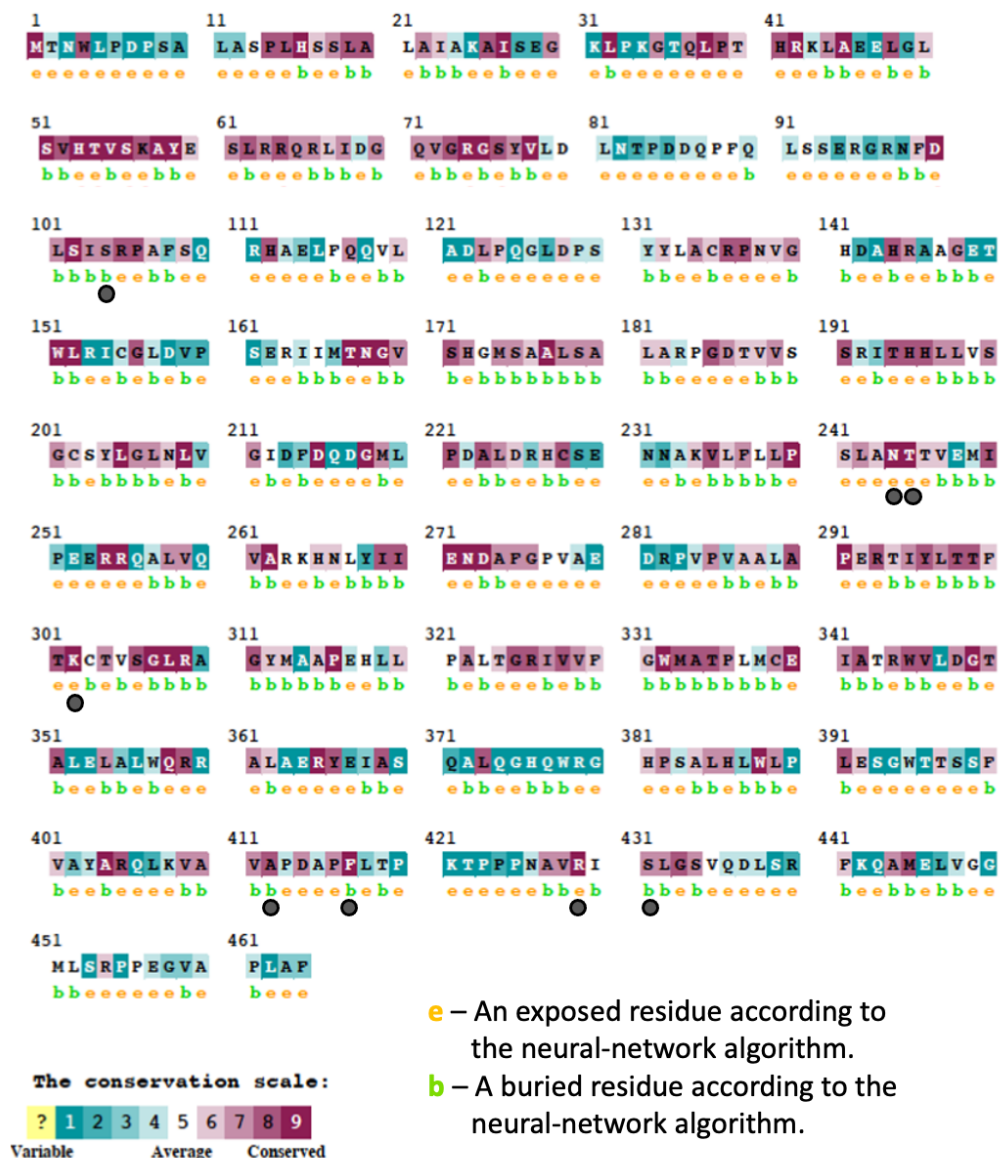
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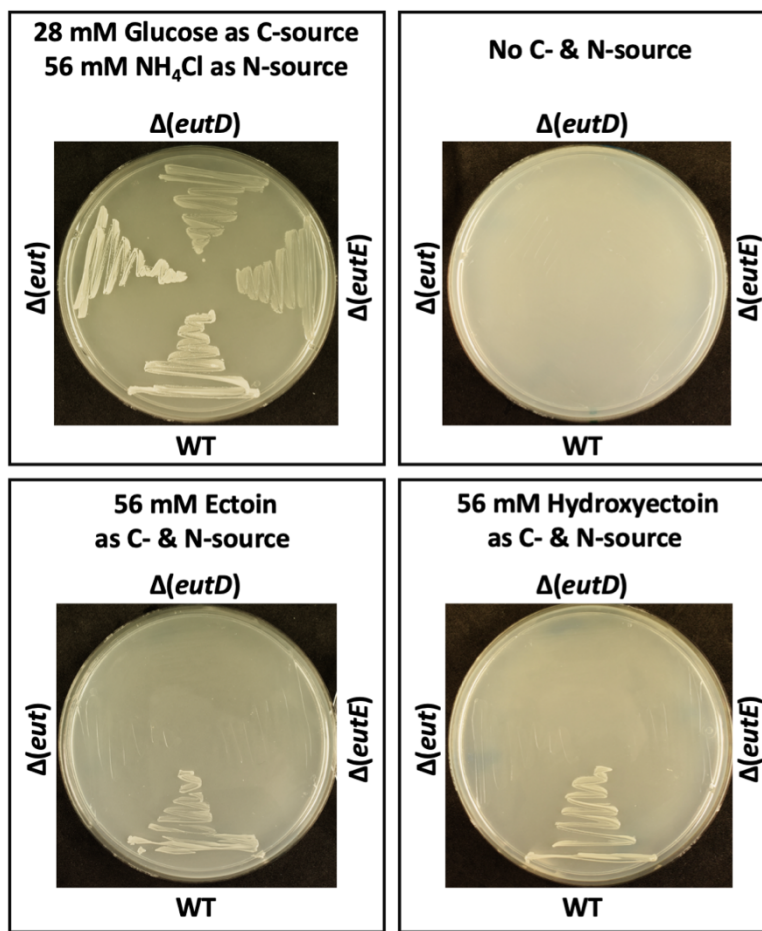
Supplementary Figure S1. Phylogenomics of EnuR-type proteins.

A phylogenetic tree of the ectoine/-5-hydroxyectoine hydrolase EutD was established using the iTol server (Letunic and Bork, 2019). The tree of 364 EutD-type proteins (Mais et al., 2020) is rooted with *Escherichia coli* and *Homo sapiens* aminopeptidases (Bradshaw et al., 1998; Wilce et al., 1998). The phylogenetic groups of microorganisms possessing EutD-type ectoine/5-hydroxyectoine hydrolases are highlighted in the two outer circles and the ectoine/5-hydroxyectoine catabolic gene clusters were analyzed for the presence of EnuR homologues in their vicinity as indicated in the inner circle.



Supplementary Figure S2. Evolutionary conservation of EnuR-type proteins.

The degree of amino acid conservation of individual amino acids of the EnuR protein of *Ruegeria pomeroyi* DSS-3 using the ConSurf server (Berezin et al., 2004). The amino acid sequences of 278 EnuR-like proteins were used in an alignment to derive a conservation matrix. The amino acid sequence of the EnuR *R. pomeroyi* DSS-3 query protein (SPO1148) (Moran et al., 2004) is displayed with the degree of evolutionary conservation at position in the protein chain. Each site is color-coded according to the degree of conservation. The first row below the sequence lists the predicted burial status of the site (see legend) (Berezin et al., 2004). Black dots mark the amino-acids determined in the *in-silico* modelling and docking experiments to be crucial for ligand-binding.



Supplementary Figure S3. Utilization of ectoines by *R. pomeroyi* DSS-3 as nutrients.

Growth of the *R. pomeroyi* wild-type strain J470 and its mutant derivatives [ASR6 $D(eut::gm^R)$, ASR8 $D(eutD::gm^R)$, and LHR7 $D(eutE::Gm^R)$] on basal minimal medium agar plates containing 28 mM ectoine or 5-hydroxyectoine when used either as sole carbon or nitrogen source. The $D(eut::gm^R)$ allele removes the entire ectoine/5-hydroxyectoine importer and catabolic gene cluster (from *enuR* to *atf*; see **Figure 1A**). Colonies were picked from basal medium agar plates containing glucose and NH₄Cl as carbon and nitrogen sources and streaked onto basal minimal agar plates containing the indicated carbon and nitrogen sources. The agar plates were incubated at 30° C for five days.

Supplementary Table S1. Phylogenetic groups of ectoine-consumers and distribution of EnuR-like proteins.

Phylogenetic group	Number of organisms	Organisms possessing EnuR like proteins	Percentage of organisms possessing EnuR [%]
<i>Rhizobiales</i>	117	103	88.0
<i>Rhodobacteraceae</i>	89	70	78.7
<i>Burkholderiales</i>	52	50	96.2
<i>Pseudomonales</i>	33	29	87.9
<i>Oceanospirillales</i>	25	5	20.0
<i>Vibrionales</i>	22	12	54.5
Minor groups	25	9	34.6
Total	363	278	76.6

Using computational tools provided via the IMG/M web-server (Chen et al., 2021), 363 microbial genome sequences (out of 8 850 inspected genome sequences) contained juxtapositioned *eutD/eutE* pairs in their ectoine/5-hydroxyectoine catabolic gene clusters (Mais et al., 2020). The taxonomic association of the corresponding microorganisms were assessed and the presence of *enuR*-type genes in the immediate vicinity of the ectoine/5-hydroxyectoine catabolic gene clusters was tabulated.

Supplementary Table S2. Docking of inducers into the presumed effector binding site of EnuR.

Inducer molecule	Binding energy (kCal/ mol)	Residues involved in hydrogen bonding
Hydroxy- α -ADABA	-7.3	Asn244, Thr245, Phe417, Ser431, Ser104
α -ADABA	-5.9	Asn244, Ser104, Ser431, Thr245
DABA	-3.8	Ser431

Results of the docking studied using AutoDock Vina (Trott and Olson, 2010) showing the predicted free energy change upon ligand-binding by the *R. pomeroiyi* DSS-3 EnuR protein, and tabulation of the amino acids predicted to be involved in inducer-binding.

Supplementary Table S3. Conservation of amino acids of EnuR predicted to be involved in α -ADABA and hydroxy- α -ADABA binding.

Residue	Conservation	Functional replacement
Ser104	274/278	3 Thr; 1 Ala
Asn244	260/278	1 Ser; 12 Gly; 5 Ala
Thr245	196/278	2 Tyr; 2 Gln; 4 Arg; 4 Asn; 57 Met; 4 Leu; 3 Lys; 2 His; 4 Phe
Lys302	278/278	
Ala412	148/278	123 Leu; 3 Met; 4 Cys
Phe417	278/278	
Arg429	278/278	
Ser431	133/278	7 Ala; 135 Cys; 3 Asn

The amino acid sequences of 278 EnuR-type proteins were aligned with Jalview (Waterhouse et al., 2009) and the conservation of those amino acids implicated by our modelling and docking studies for the binding of the inducer molecules α -ADABA and hydroxy- α -ADABA were assessed.

Supplementary Table S4. Strains used in this study.

Strain	Genotype or description	Source or reference
<i>Escherichia coli</i> DH5a	Used for routine cloning purposes	Invitrogen, Karlsruhe, Germany
<i>Escherichia coli</i> BL21 (DE3)	Strain, used for overexpression	Stratagene, La Jolla, CA
<i>Ruegeria pomeroyi</i> DSS-3	Wild-type strain	(Moran et al., 2004)
<i>Ruegeria pomeroyi</i> J470 ^a	Rif ^R derivative of the wild-type strain	(Todd et al., 2012)
<i>Ruegeria pomeroyi</i> ASR6 ^b	<i>R. pomeroyi</i> J470 $\Delta(enuR-atf::gm)1$	(Schulz et al., 2017a)
<i>Ruegeria pomeroyi</i> ASR8 ^b	<i>R. pomeroyi</i> J470 $\Delta(eutD::gm)1$	(Schulz et al., 2017a)
<i>Ruegeria pomeroyi</i> LHR7 ^b	<i>R. pomeroyi</i> J470 $\Delta(eutE::gm)1$	This study

^aRif^R: Resistant against the antibiotic rifampicin.

^b*gm*: Genetic determinant conferring resistance against the antibiotic gentamycin.

Supplementary Table S5. Plasmids used in this study.

Plasmid	Genotype or description	Source or reference
pRK2013 ^a	Helper plasmid for tri-parental mating, Kan ^R	(Figurski and Helinski, 1979)
pK18mobsacB	Suicide vector for <i>R. pomeroyi</i> , Kan ^R	(Kvitko and Collmer, 2011)
p34S-gm ^c	Plasmid carrying a gentamicin (Gm ^R) resistance cassette	(Dennis and Zylstra, 1998)
pLH73	pK18mobsacB with flanking regions of the <i>eutE</i> -gene, interrupted with a Gm ^R cassette, Kan ^R	This study
pEntry51	Cloning vector for IBA-Stargate cloning	IBA GmbH, Göttingen, Germany
pASG-IBA3	<i>E. coli</i> expression vector carrying a TetR-controlled and anhydrotetracyclin-responsive <i>tet</i> promoter	IBA GmbH, Göttingen, Germany
pBAS3 ^d	pASG-IBA3 with synthetic, codon optimized <i>enuR</i> gene	(Schulz et al., 2017b)
pBAS17 ^d	pBAS3 with codon exchange mutation (AAA/CAT) in the codon-optimized <i>enuR</i> leading to the replacement of Lys-302 with a His residue	(Schulz et al., 2017a)
pLH17 ^d	pASG-IBA3 with synthetic, codon optimized sole aminotransferase domain of the <i>enuR</i> gene	This study
pLH26 ^d	pLH17 with codon exchange mutation (AAA/CAT) in the codon-optimized <i>enuR</i> gene leading to the replacement of Lys-302 with a His residue	This study

^aKan^R: Resistant against the antibiotic kanamycin.

^bgm: Genetic determinant conferring resistance against the antibiotic gentamycin.

^cThe *R. pomeroyi* DSS-3 *enuR* gene (or a segment thereof) carried by these plasmids was codon-optimized for enhanced expression in *E. coli* (Schulz et al., 2017b). The DNA-sequence of this synthetic gene is available in GenBank under accession number KU891821.

Supplementary Table S6. Oligonucleotides used in this study.

Primer	Sequence	Reference, description
ATD_pEntry_fw	AAGCTCTTCAATGCGTAATTTTGATCTGAGCA TTAGCCG	IBA-Stargate cloning of the ATD of <i>Ruegeria pomeroyi</i> EnuR
ATD_pEntry_rev	AAGCGGCTCTTCTCCCAAGCTCTTACCCAAA TGCCAG	IBA-Stargate cloning of the ATD of <i>Ruegeria pomeroyi</i> EnuR
L263_fw	GGTCGGCGGCATGCTG	EMSA-fragments for the <i>uehA</i> - operator region
L229_rev_dye	GGTTTCCTCCCAAATGTCATGGG	EMSA-fragments for the <i>uehA</i> - operator region
Δ eutE_F1_fw	ACAGCTATGACATGATTACGCGCATCTGACCT GGGACGAT	Construction of plasmid pLH73
Δ eutE_F1_rev	ttcgagctcgAGTCCTTACGAACATCTTGCGCG G	Construction of plasmid pLH73
Δ eutE_gm_fw	GTGAAGGACTcgagctcgaattgacataagcctggt	Construction of plasmid pLH73
Δ eutE_gm_rev	GGTCCGCCTCtgttaggtggcggtacttgggt	Construction of plasmid pLH73
Δ eutE_F2_fw	ccacctaacaGAGGCGGACCCATGCA	Construction of plasmid pLH73
Δ eutE_F2_rev	ATCCCCGGGTACCGAGCTCGGCTGGCGCCGT CACT	Construction of plasmid pLH73
MST BS WT fw	TAACATTGTCGCGGACAATAAAAAAATTGA CATGCAGTACAATTCCC	Fragment for MST ^a
MST BS WT rev	GGGAATTGTAAGTGCATGTCAATTTTTTTATTG TCGCGGACAATGTTA	Fragment for MST ^a

^aMST: microscale thermophoresis

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